PHLOROTANNINS OF PHAEOPHYCEA LAMINARIA OCHROLEUCA*

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Abstract—From the mixture of acetylated phenols of Laminaria ochroleuca several fractions consisting of one or more components were isolated and analysed. Several substances, new or known from other seaweeds were identified: tetraphlorethol-A-nonacetate, fucophlorethol-B-octacetate, fucodiphlorethol-C-decacetate, pentafuhaloltridecacetate, and heptafuhaloloctadecacetate. A structure for an isomer of tetraphlorethol-A-nonacetate named tetraphlorethol-B-nonacetate was suggested. Additional phlorotannins were shown to be present, for which only partial structures could be proven.

INTRODUCTION

Recently we described the isolation of acetylated di- and triphlorethol C together with their monochlorinated derivatives from the Phaeophycea Laminaria ochroleuca [1,2]. In the enriched acetylated extract further low molecular weight phlorotannins are present. Definite structures for some of them were determined, and others were at least partially characterized.

RESULTS AND DISCUSSION

A spot at R_c 0.37 on TLC (Si gel 60, CHCl₃-Me₂CO, 47:3) contained a mixture of compounds 2 and 3. These compounds were eluted as a mixture from a HPLC Si gel column with various gradient programs (CHCl₃-EtOH). In the electron impact-induced MS, 2 had an M⁺ at m/e 876 (corresponding to $C_{42}H_{36}O_{21}$) and lost ketene (42 mu) up to nine times to yield the free phenol C₂₄H₁₈O₁₂ (m/e 498). Compound 2 therefore seemed to be a member of the phlorethol series with four benzene rings and nine acetylated HO groups. In many respects, the ¹H NMR spectrum was very similar to that of triphlorethol-C-acetate (1) [1] with resonances at δ 6.95, 2.26, 2.08 and 2.06 due to two terminal 2,4,6triacetoxylated phenoxyl groups and additional resonances for three isochronic acetoxy protons at 2.20 and an AB₂ spin system between 6.31 and 6.45 with $J_{AB} \sim 2 \text{ Hz}$. Because of the low amount of isolated material, the spectra had to be taken by the CAT-technique. Therefore, the line width was large and the signals were not resolved totally. In the spectra of 2 additional resonances beyond 1 are found at 6.71 (2 aromatic H) and 2.12 (6 acetoxy-H). They must belong to a diacetoxybenzene ring linked by two para-situated ether bridges. Thus, compound 2 was 2,6,5'-triacetoxy-4,3'-bis(2,4,6-

$$1 R_1/R_2 = OAc$$

$$\mathbf{2} \quad \mathbf{R}_1 = \mathbf{OAc} \qquad \mathbf{R}_2 = -\mathbf{O} - \underbrace{\mathbf{A'}}_{\mathbf{OAc}} - \mathbf{OAc}$$

$$\mathbf{A} \quad \mathbf{R}_1 = -\mathbf{O} - \mathbf{O} - \mathbf{R}_2 = \mathbf{O} - \mathbf{R}_2$$

triacetoxyphenoxy) diphenylether, commonly called tetraphlorethol-A-nonacetate. As mentioned above, 2 was contaminated by small quantities of another substance, 3. Up to now, the mixture could not be resolved either by TLC or by HPLC. However, the MS and the ¹H NMR resonances of 3 were identical with those of fucophlorethol-B-octacetate recently isolated from Cystoseira baccata [3].

Compound 4, combined with an approximate 10% impurity of 5, was found at R_f 0.33. Substance 4 showed a molecular ion at m/e 876 which eliminated up to nine

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3
$$R_1 = OAc$$
: $R_2 = -O$

5
$$R_1 = [OPh(OAc)_2]_2OAc$$
 $R_2 = -1$

6
$$R_1 = OAc$$
 $R_2 = -O$

OAc

OAc

OAc

ketene units. It consisted of four ether-linked benzene rings substituted by nine acetoxy groups, therefore 4 appeared to be an isomer of 2. Compound 4 showed the same ¹H NMR resonances for the rings A, B, respectively A', as 1 or 2. The AB system of ring C ($v_A = 6.68$, $v_B =$ 6.71 ppm, $J_{AB} = 2 \text{ Hz}$) was overlapped by some signals of 5. As the Me signals around 2 ppm cannot be assigned exactly, it is impossible to absolutely prove the existence of a 2,3-diphenoxy-1,4-diacetoxybenzene structure element for ring C. Such an element would be in agreement with the theory of biosynthesis. It should be pointed out that Glombitza et al. [4] demonstrated the presence of the same substitution pattern in a desacetoxyheptafuhalol with different ¹H NMR values ($v_A = 6.51$, $v_B =$ 6.72 ppm). Attempts to synthesize a model substance with a similar substitution pattern have failed up to now. Tentatively, 4 is thought to have the structure of a tetraphlorethol-B-nonacetate isomeric to 2.

A signal at m/e 912 was attributed to the molecular ion of 5. The elimination of H_2O from partially deacetylated fragments containing three or four rings leading to ions with a dibenzofuran structure [5], together with the ¹H NMR spectrum, gives evidence that 5 is a fucodiphloretholdecacetate. The resonances at δ 6.99, 6.70 (each 2 aromatic H) and 2.04, 1.97 (each 6 isochronic acetoxy protons) were typical signals of a 4'-phenoxy substituted 2,4,6,2',6'-pentacetoxybiphenyl moiety. Partial structures like this have been described for some of the phlorotannins found in Fucus vesiculosus [5]. As yet, nothing can be said about the two other rings of 5.

From a spot at R_f 0.27, a MS was taken with a molecular ion at m/e 918 ($C_{44}H_{38}O_{22}$, 6). A free phenol

 $(m/e 498, C_{24}H_{18}O_{12})$ was left after a ten-fold ketene elimination. Some of the partially or totally deacetylated ions lost H_2O . This behaviour is typical of the abovementioned fucophlorethols. Resonances at δ 7.09 (1 H), 7.0 (2 H), 2.27 (3 H), 2.06 (9 H), 2.03 (3 H) and 1.86 (3 H) gave evidence that 6 contained a 2,4,6,2',4',6'-hexacetoxy-biphenyl-3-phenoxy element like 3. Thus, 6 seemed to be an isomer of 5. Between δ 6.29 and 6.46, 6 showed a poorly resolved three-spin coupling system similar to 1. A distinct signal at 6.95 and further resonances in the acetoxy-range showed the presence of a symmetrically substituted 2,4,6-triacetoxyphenoxy group. According to the spectral data, 6 had the structure of a 2,4,6,2',4',6'-hexacetoxy-3-[5-{2,4,6-triacetoxyphenoxy}-3-acetoxyphenoxy] biphenyl (fucodiphlorethol-C-decacetate).

The spot at R_f 0.26 was an inseparable phlorotannin mixture. One component, 7, gave a molecular ion at m/e 1126 ($C_{54}H_{46}O_{27}$) which fragmented by successive loss of ketene to an ion at m/e 748. Compound 7 seemed to be the next higher homologue of either 6 or 5. The ¹H NMR spectrum did not allow further conclusions as to the structure of 7.

Laminaria ochroleuca specimens from two different locations, Roscoff/Le Loup and Ménéham, were investigated. The Ménéham material contained two more derivatives (8 and 9) of the fuhalolacetate type. Compound 8 was detected on the TLC at R_{ℓ} 0.28 and 0.8 mg was isolated. The MS and ¹H NMR spectrum were taken. Compound 8 proved to be identical with a pentafuhaloltridecacetate previously isolated from Halidrys siliquosa [4] and Bifurcaria bifurcata [6], the structure of which was confirmed by 13C NMR spectroscopy. Substance 9 was isolated from a spot at $R_c 0.15$. The MS showed fragmentation patterns that varied distinctly according to the operation conditions. Electron impact energy of 70 eV and temperatures up to 240° resulted in fragments with 2-5 rings and of a low degree of hydroxylation in agreement with previous results [4, 7]. When the temperature was increased to 280° and the ionization to 100 eV, ions of the ketene elimination series, starting from the molecular ion, and daughter ions consisting of 3 or 5 phloroglucinol units with three additional HO groups became visible. These 3 and 5 rings originated from the central part of the molecule. Further temperature increase resulted in catalytic dehydrogenation reactions, which led to series differing from the main series by -2H or -4H. Possibly derivatives of o-quinones, dibenzodioxines, or both are formed. A sophisticated analysis of the fragments seemed establish the structure proposal published recently [4, 7] for heptafuhaloloctadecacetate.

No further detailed analysis of additional fractions consisting of mixtures of 4- to 7-ring compounds was possible. However, partial structures for some could be derived from the spectral data. They contained only members of the phlorethol (ether-linked phloroglucinols) or fucophlorethol series (phenoxy substituted biphenyls with phloroglucinol pattern). Some of these compounds are chlorinated.

EXPERIMENTAL

Extraction and chromatography. 15 kg lyophilized, pulverized thalli from Roscoff/Brittany (April 1974) and 10 kg from Ménéham/Brittany (May 1973) were extracted as described in ref. [1]. The phenolic fraction was enriched and prefractionated. Separation was carried out by combination of TLC and HPLC.

For PLC, Si gel plates (Merck 60, F_{254} Fertigplatten, 0.25 mm) were used with different solvent systems (mainly CHCl₃-Me₂CO, 47:3; CCl₄-CHCl₃, 4:1) and repeated development. Gradient elution systems with various CHCl₃-EtOH mixtures (0.2-5% EtOH) were used with a Si gel HPLC column (Partisil 10, 25 cm, 9 mm ϕ). Detection by UV absorption at 275 nm. All given R_f values are in reference to Si gel plates (Merck 60, F_{254} , 0.25 mm) with the solvent system CHCl₃-Me₂CO (47:3).

Tetraphlorethol-A-nonacetate (2). 5th zone of TLC, R_f 0.37. 2.6,5'-triacetoxy-4,3'-bis (2,4,6-triacetoxyphenoxy) diphenylether, 6.5 mg/15 kg algae; ¹H NMR (CDCl₃, 100 MHz, CAT): δ 6.95 (4H), 6.71 (2H), 6.45-6.31 (3 spin coupling system, $J_{AB} \sim 2$ Hz), 2.26 (6H), 2.20 (3H), 2.12, 2.08, 2.06 (each 6H).

Fucophlorethol-A-nonacetate (3). 2,4,6,2',4',6'-Hexacetoxy-3-(3,5-diacetoxyphenoxy)biphenyl, 5th zone of TLC, R_f 0.37; in a ratio of 1:7 together with 2; corresponding to 1 mg/15 kg algae; ¹H NMR and MS values were identical with the described values in [3].

Tetraphlorethol-B-nonacetate (4). Tentatively identified as 3,1',5'-triacetoxy-1,3'-bis(2,4,6-triacetoxyphenoxy)diphenylether; 7th zone of the TLC, R_f 0.33; 8 mg/15 kg algae; ¹H NMR (CDCl₃, 100 MHz, CAT): δ 6.94 (4 H), AB system (2 H, v_A = 6.46, v_B = 6.33, J_{AB} = 2.1 Hz), 2.26, 2.20, 2.07, 2.06 (in a ratio of 2.5:1.5:4:0.8).

Fucodiphloretholdecacetate (5). 7th zone of the TLC, R_f 0.33; in a ratio of 1:10 together with 4, corresponding to 1 mg/15 kg algae.

Fucodiphlorethol-C-decacetate (6). 2,4,6,2',4',6'-hexacetoxy-3-[5-(2,4,6-triacetoxyphenoxy)-3-acetoxyphenoxy] biphenyl; 10th zone of the TLC, R_f 0.27; 2.7 mg/15 kg algae; ¹H NMR (CDCl₃, 90 MHz): δ 7.09 (1 H), 7.0, 6.95 (each 2 H), 6.45-6.29 (3 spin coupling system, $J_{AB} \simeq 2$ Hz), 2.27 (6 H), 2.20 (3 H), 2.10 (6 H), 2.06 (9 H), 2.03, 1.86 (each 3 H).

Fucotriphloretholdodecacetate (7). 11th zone of the TLC, $R_f 0.26$.

Pentajuhaloltridecacetate (8). 9th zone of the TLC, R_f 0.28; 0.8 mg/10 kg algae, ¹H NMR and MS values are identical with those in refs. [4] and [6].

Heptafuhaloloctadecacetate (9). $R_7 0.15$; 43 mg/10 kg algae; IR. MS (70 eV), UV, ¹H NMR (CDCl₃ and Me₂CO- d_0) are identical with the description in refs. [4] and [7]. MS 100 eV, temp. >280°): ketene elimination starting and ending with: m/e 430-262, 432-264, 532-280, 530-278, 680-386, 682-388, 698-404, 696-402, 756-420, 712-628, 1090-712, 948-738, 1030-736, 1130-752, 1070-944, 1212-960.

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REFERENCES

- Glombitza, K. -W., Koch, M. and Eckhardt, G. (1976) Phytochemistry 15, 1082.
- Glombitza, K. -W., Koch, M. and Eckhardt, G. (1977) Phytochemistry 16, 796.
- Glombitza, K. -W., Wiedenfeld G. and Eckhardt, G. (1978) Arch. Pharm. 311, 393.
- Glombitza, K. -W., Geisler, C. and Eckhardt, G. (1980) (in preparation).
- Glombitza, K.-W., Rauwald, H.-W. and Eckhardt, G. (1977) Planta Med. 32, 33.
- Glombitza, K. -W., Rösener, H. U. and Koch, M. (1976) Phytochemistry 15, 1279.
- Sattler, E., Glombitza, K.-W., Wehrli, F. W. and Eckhardt, G. (1977) Tetrahedron 33, 1239.